

## **Cambridge International Examinations** Cambridge International Advanced Subsidiary and Advanced Level

## BIOLOGY

9700/52 October/November 2016

Paper 5 Planning, Analysis and Evaluation MARK SCHEME Maximum Mark: 30

Published

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	Page 2	Mark Scheme	Syllabus	Paper
ſ		Cambridge International AS/A Level – October/November 2016	9700	52

Question	Answer	Mark	Additional Guidance
1(a)(i)	<i>independent:</i> <u>concentration</u> of potassium chloride/KCl;	2	A different concentrations of potassium chloride
	dependent: number of stomata open/closed;		A number open and closed
1(a)(ii)	three from:	3	<b>A</b> volumes either in descriptions or a table
	correct volumes of water and KC <i>l</i> solution for making <u>all</u> four dilutions with units ;;		max 1 for correct volumes making 1, 2 or 3 dilutions
	method of measuring volumes ;		
	ref. to stirring/mixing;		
1(b)(i)	<i>idea of:</i> the higher the concentration of (potassium chloride/KC <i>l</i> ) the greater the number of stomata open/ <b>ora</b> <b>or</b>	1	<ul> <li><i>R</i> in terms of degree/speed of opening and closing of stomata e.g. more KC1 the stomata are wider.</li> <li>A a null hypothesis:</li> </ul>
	the higher the concentration of (potassium chloride/KCl) the lower the number of stomata open/ <b>ora</b>		
	<b>or</b> the number of open stomata is directly proportional/inversely proportional to the concentration of potassium chloride/ <b>ora</b> ;		

Page 3	Mark Scheme	Syllabus	Paper
	Cambridge International AS/A Level – October/November 2016	9700	52

Question	Answer	Mark	Additional Guidance
1(b)(ii)	<ul> <li><i>five from:</i></li> <li>1 <i>ref. to</i> putting the strips into (all KC<i>l</i>) solutions in appropriate containers;</li> <li>2 <i>ref. to</i> keeping in the dark (when in solution);</li> </ul>	5	e.g. beakers, watch glasses, Petri dishes <b>R</b> test-tubes/boiling tubes/cavity slides
	<ul> <li>3 <i>ref. to</i> mounting on a slide <b>and</b> using a (light) microscope (to count/observe the number of stomata);</li> <li>4 <i>ref. to</i> count/record the number of stomata that are open or</li> </ul>		R electron/electronic microscope/hand lens/magnifying glass
	<ul> <li>closed ;</li> <li><i>ref. to</i> a method standardising the counting open/closed stomata ;</li> <li><i>ref. to</i> making several counts on each leaf strip <b>and</b> taking a <u>mean</u>/to identify anomalies ;</li> </ul>		<ul> <li>e.g. out of the same fixed number of stomata</li> <li>or in field of view (at the same magnification)</li> <li>A a minimum of 3 counts on one strip</li> <li>I ref. to repeating whole experiment three times</li> </ul>
	<ul> <li><i>control variables max 2 (7–9)</i></li> <li><i>ref. to</i> using suitable equipment for cutting <b>and</b> measuring strips (of same length and width/size/area);</li> </ul>		R metre rule
	<ul> <li>8 <i>ref. to</i> a method of maintaining a constant temperature ;</li> <li>9 covering to prevent evaporation ;</li> </ul>		A incubator/temperature controlled room/water-bath if appropriate to apparatus
	10 one of: ref. to low risk ; examples of hazard and precaution ;		<b>R</b> no risk

Page 4	Mark Scheme	Syllabus	Paper
	Cambridge International AS/A Level – October/November 2016	9700	52

Question	Answer													Mark		Additional Guidance				
1(c)(i) <i>ref. to</i> using (eyepiece) graticule to me										asur	sure (the aperture) ;								2	R if use graticule and stage micrometer to measure
one from calibrating the (eyepiece) graticule with a (stage) micrometer AW ; convert/calibrate the eye piece units to $\mu m/mm$ ;												<b>A</b> <i>ref. to</i> converting eyepiece units using conversion/calibration factor								
1(c)(ii)	two (for one mark) from															1				
	time mir	me / stomatal aperture / µm																		
	0	0.5	0.1	0.2	0.3	0.4	0.1	0.5	0.2	0.3	0.3	0.1	0.2	0.2	0.2	0.4				
	60	0.9	1.1	1.0	1.3	1.2	1.8	1.5	0.8	0.2	1.3	1.1	0.8	1.0	1.9	0.9				
	120	1.9	2.4	2.6	2.6	2.5	2.2	2.8	2.4	2.4	3.9	2.6	2.3	2.5	2.2	2.7				
	180	4.1	4.8	4.2	4.0	5.7	4.7	3.9	4.1	5.5	4.5	4.3	4.0	3.1	4.1	4.3				
1(c)(iii)	0.03	<u>5</u> ;																	1	

Page 5	Mark Scheme	Syllabus	Paper
	Cambridge International AS/A Level – October/November 2016	9700	52

Question	Answer	Mark	Additional Guidance
1(c)(iv)	<i>three from</i> measure more stomata/all the stomata (per epidermal strip) ; select stomata to be measured randomly ;	3	if specify a number, should be 10 or more
	use more leaves/epidermal strips ; measure at shorter (time) intervals/more frequently ;		<b>R</b> use different types of plant
1(d)	<i>idea that</i> the longer the time of light exposure the wider stomata open / the wider the aperture ;	1	R idea of different light intensity
	Tota	: 19	

Question	Answer	Mark	Additional Guidance
2(a)	<i>two (for one mark) from</i> number of fields studied ; (width of) the headland/strip ; (type of) cereal/crop ;	1	A length <i>if qualified by</i> 6 <i>m</i>
2(b)(i)	data is nominal/categoric or testing the difference between observed (O) and expected (E) results ;	1	A data can be grouped/is discrete
2(b)(ii)	there is no <u>significant difference</u> between number of butterflies of each species when headland sprayed and when not sprayed ;	1	A without herbicide/not treated/control for not sprayed A with herbicide/treated for sprayed

Page 6	Mark Scheme	Syllabus	Paper
	Cambridge International AS/A Level – October/November 2016	9700	52

Question	Answer	Mark	Additional Guidance
2(b)(iii)	species <b>Q</b> O E $(O-E)^2$ $\frac{(O-E)^2}{E}$	3	if E is correct, but one row is processed incorrectly, allow ecf for correct addition to obtain $\chi^2$ value
	number on headland 3 20 289 14.45 ;		max 2
	number on headland not sprayed with herbicide 37 20 289 14.45 ;		
	$\chi^2 = 28.9$ ;		
2(b)(iv)	<u>3.84</u> ;	1	
2(b)(v)	significant (at p<0.001)/herbicide is causing the number of butterflies to decrease ;	1	ecf from errors in (iii) and/or (iv)
2(c)	<ul> <li>three from <ol> <li>idea that where herbicide has been used there are fewer/smaller population of all species investigated;</li> <li>idea of (decrease/difference) in species S is only one that is not significant/ora;</li> <li>herbicide has greatest effect on the population of R (and Q);</li> <li>ref. to the sequence of the severity of the effect of the herbicide;</li> <li>probability of the results being due to chance is less than 5% for all species except S (and Q);</li> </ol> </li> </ul>	3	sequence is ( <b>R</b> >)V/W>T/U>S <i>if <b>R</b> included in the sequence allow mp3 and mp4</i> <b>A</b> probability of the result being due to herbicide is more than 95% for all species except <b>S</b>
	Total:	11	