

UNIVERSITY OF CAMBRIDGE INTERNATIONAL EXAMINATIONS General Certificate of Education Advanced Subsidiary Level and Advanced Level

CANDIDATE NAME					
CENTRE NUMBER			CANDIDATE NUMBER		

167082794

BIOLOGY 9700/31

Paper 3 Advanced Practical Skills

May/June 2008

2 hours

Candidates answer on the Question Paper.

Additional Materials: As listed in the Instructions to Supervisors

READ THESE INSTRUCTIONS FIRST

Write your Centre number, candidate number and name on all the work you hand in.

Write in dark blue or black pen.

You may use a soft pencil for any diagrams, graphs or rough working.

Do **not** use staples, paper clips, highlighters, glue or correction fluid.

DO NOT WRITE IN ANY BARCODES.

Answer all questions.

At the end of the examination, fasten all your work securely together.

The number of marks is given in brackets [] at the end of each question or part question.

You are advised to spend one hour on each question.

The length of smallest diving stage micro	ision on the
For Exam	iner's Use
1	
2	
Total	

This document consists of 10 printed pages and 2 blank pages.



You are reminded that you have only one hour for each question in the practical examination. You should read carefully through the whole of each question and then plan your use of the time to make sure that you finish all of the work that you would like to do.

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- 1 You are required to carry out tests, using only the reagents provided, to identify each of the solutions **S1**, **S2** and **S3**.
 - One of the solutions is glucose, another a protein and the third a carbohydrate other than glucose.

You are required to identify each of the solutions, **S1**, **S2** and **S3**. You must use only the reagents provided.

(a) (i) Prepare and use the space below to record the test used, observations and conclusions.

[5]

•••••			
			[3]
		ation in Table 1.1 to estimate th	ne concentration o
the glucose			
		Гable 1.1	
	colour	glucose concentration / mol dm ⁻³	
	blue	0.00	
	green	0.01	
	yellow	0.05	
	red	0.10	
concontratio	on of alucoso solution	١	[4]
dentify two	sources of error in e	stimating the concentration of th	ne solution.
1			

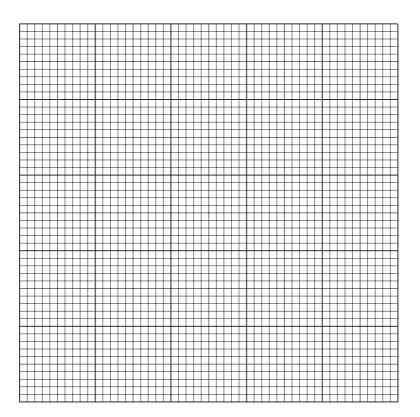
eplicates were run,	starting wi	th fresh ma	terials ea	ch run.		
data in Table 1.2 we	ere obtaine	d.				
data in Table 1.2 we	ere obtaine	d. Table 1.2				
percentage	ere obtaine	Table 1.2	ssion of li	ght/arbitrai	y units	
	first run	Table 1.2	ssion of li third run	ght/arbitrai fourth run	ry units fifth run	mean
percentage concentration of	first	transmis second	third	fourth	fifth	mean 92
percentage concentration of starch suspension	first run	transmis second run	third run	fourth run	fifth run	
percentage concentration of starch suspension 0.0	first run 92	transmis second run 91	third run 92	fourth run 94	fifth run 89	92
percentage concentration of starch suspension 0.0 0.5	first run 92 61	transmin second run 91 60	third run 92 59	fourth run 94 60	fifth run 89 58	92
percentage concentration of starch suspension 0.0 0.5	first run 92 61 41	transmis second run 91 60 41	third run 92 59 42	fourth run 94 60 43	fifth run 89 58 41	92

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(b)

(iii)	Plot	а	graph	of	percentage	concentration	of	starch	suspension	against	the
	transmission of light using the student's results.										

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[3]

(c) The student's hypothesis was:

Transmission of light is proportional to the concentration of starch suspension.

Draw an appropriate conclusion to the student's experiment. You should include in your conclusion whether the experimental data support the hypothesis and produce a revised hypothesis, if necessary.

.....[2

[Total : 21]

2 J1 is a slide of a stained transverse section through the leaf of a xerophyte.

You are also provided with an eyepiece graticule that has been fitted to the eyepiece of your microscope and a stage micrometer scale.

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(a) (i) Draw a large low-power plan diagram of a part of **J1** as shown in **Fig. 2.1**. Labels are not required.

drawing of this part required



[4]

(ii) Fig. 2.2 is a photomicrograph of part of J1.

Examiner's tip of Use

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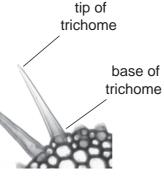


Fig. 2.2

Carefully examine a similar area of **J1** using the high-power of your microscope.

Put a ring round the number written on the objective lens.

1/6" other ×40 4mm Identify a trichome (hair). Count the number of eyepiece graticule divisions across the width of the base of the trichome.

Remove the slide **J1** and replace it with the stage micrometer scale.

number of eyepiece graticule divisions

Using the same magnification, adjust the focus until you can see the eyepiece graticule on top of the stage scale.

Count the number of eyepiece graticule divisions that match an exact number of stage scale divisions.

number of eyepiece graticule divisions

number of stage micrometer scale divisions

Use this information to calculate the actual width of the trichome on your slide **J1**.

Show your working.

actual width of trichomeµm

[4]

(iii)	Suggest how an error in measuring the trichome could occur.	For Examiner
		Use
	[1]	
(iv)	Suggest the purpose of the trichomes on the leaf of the xerophyte.	
	[1]	

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(b) In the space below, make a large high-power drawing of three cells from the inner layer (at **X**) and three cells from the outer layer (at **Y**) of **J1**, as shown in **Fig. 2.3**.

For Examiner's Use

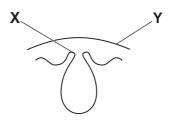


Fig. 2.3

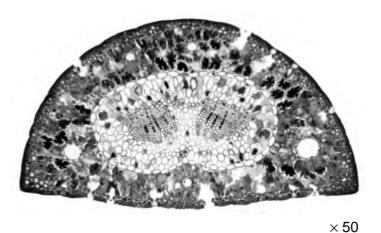
three cells from the inner layer (at X)

three cells from the outer layer (at Y)

[4]

(c) Fig. 2.4 is a photomicrograph of a transverse section of a leaf of another xerophyte.

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. .

Fig. 2.4

Prepare the space below so that it is suitable for you to compare and contrast the section on slide **J1**, with the section shown in **Fig. 2.4**.

Record your **observations** in the space that you have prepared.

[5]

[Total: 19]

[Paper total: 40]

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